User Manual

FAVORGEN

FavorPrep[™] 96-Well Viral DNA/RNA Kit

- For 96-well high-throughput extraction of viral nucleic acid from cell free samples such as serum, plasma, body fluids and the supernatants of cell cultures

Kit contents:

Cat. No.: (Q'ty)	FAVRE 96001 (1 plate)	FAVRE 96002 (2 plates)	FAVRE 96004 (4 plates)
VNE Buffer	60 ml	120 ml	120 ml × 2
AD Buffer 🔺	5 ml	10 ml	10 ml × 2
Wash Buffer 1 (concentrate)	55 ml	110 ml	110 ml × 2
Wash Buffer 2	25 ml	50 ml	50 ml × 2
RNase-free water	15 ml	30 ml	30 ml × 2
Filter Plate (96-Well DNA/RNA Binding plate)	1 plate	2 plates	4 plates
Collection Plate (96-Well 2 ml Plate)	3 plates	6 plates	12 plates
Elution Plate (96-Well PCR plate)	1 plate	2 plates	4 plates
Adhesive Film	2 pcs	4 pcs	8 pcs

▲, ■, □: Add ethanol (96~100%) to AD Buffer, Wash Buffer 1 and Wash Buffer 2 when first use. See Working Buffer Preparation.

Storage:

All components of FavorPrep[™] 96-Well Viral DNA/RNA Kit should be stored at room temperature (15~25°C).

Quality control

The quality of FavorPrep[™] 96-Well Viral DNA/RNA Kit is tested on a lot-to-lot basis according to ISO quality management system. The purified nucleic acid is checked by real-time PCR and capillary electrophoresis.

Product description:

FavorPrep™ 96-well Viral DNA/RNA Mini Kit is an excellent tool for 96-well high-through put extraction of high pure viral nucleic acid from viral cell free specimen such as, serum, plasma, body fluid and cell cultured supernatant, and from transport medium of swabs. The extraction method is based on the silica membrane/chaotropic salt technology, and the procedure involves lysis of virus, optimization of binding condition being able to make the viral nucleic acid efficiently to silica membrane, washing silica membranes to remove contaminations including salts, metabolites, nucleases and other components of body fluid, finally elution of the viral nucleic acid from the silica membrane. Compare with other harmful and time-consuming method, such as phenol/chloroform extraction and ethanol precipitation, FavorPrep™ 96-well Viral DNA/RNA Kit makes extraction of high-purity viral nucleic acid reliable, and that shortens the handling time less than 60 minutes for a 96 - preparations

Specification:

Principle: Filter Plate (glass fiber membrane) Sample size: 200 µl of serum, plasma, body fluids and the supernatant of cell cultures Plate applicability: vacuum or centrifugation Operation time: within 1 hour/96 preparations RNA Binding capacity: up to 60 µg/well Elution volume: 50~75 µl

Additional materials required

For All Protocol:

- Pipets and pipet tips, sterile
- 96~100% nuclease-free ethanol

For vacuum processing:

- A centrifugator is required for the clarification of lysate and for the alternative of elution step, capable of 5,600~6,000 X g, with a swing-bucket rotor and the adaptor for 96-well plates. • A vacuum manifold for 96-well plate and a vacuum source
- reached to -12 inches Hg are required.

For centrifuge processing:

• A centrifugator is required, capable of 5,600~6,000 X g, with a swing-bucket rotor and the adaptor for 96-well plates.

Working Buffer Preparations:

Preparation of AD Buffer A, Wash Buffer 1 and Wash Buffer 2 🗆

Add required ethanol (96~100%) to AD Buffer, Wash Buffer 1 and Wash Buffer 2 as the table indicated. Store the ethanol added AD Buffer, Wash Buffer 1 and Wash Buffer 2 at 15~25°C.

Cat. No.:	FAVRE 96001 (1 plate)	FAVRE 96002 (2 plates)	FAVRE 96004 (4 plates)
▲ Ethanol volume for AD Buffer	40 ml	80 ml	80 ml
Ethanol volume for Wash Buffer 1	10 ml	20 ml	20 ml
Ethanol volume for Wash Buffer 2	100 ml	200 ml	200 ml

Important notes:

1. Notes for sample preparation:

- Make sure everything is RNase-free when handling this system. • Buffers provided in this system contain irritants. Wear gloves and
- lab coat when handling these buffers.
- Do not thaw the frozen plasma or serum samples more than once.
- Centrifuge the plasma or serum samples at 6,000 x g for 3 minutes. If precipitates are visible. Then transfer the cleared supernatant to a new vial and processed immediately.

2. Notes for Buffers:

- Add required ethanol (96~100%) to AD Buffer, Wash Buffer 1 and Wash Buffer 2 before use. See Working Buffer Preparation.
- For handling the buffers safely please read safety Information before starting the procedure.

3. Notes for centrifuging and vacuum:

- Ensure that centrifugation speed is according to instruction of individual step.
- When using of vacuum to proceed "DNA/RNA Binding" and "Wash the Filter Plate", ensure the vacuum pressure being capable to reach to -12 inches Hg.



Brief procedure:





Plate)

Safety Information:

- 1. VNE Buffer and Wash Buffer 1 provided in this system contain irritants. Wear gloves and lab coat when handling these buffers.
- CAUTION: VNE Buffers and Wash Buffer 1 contains guanidinium salts which can form highly reactive compounds when c ombined with bleach. DO NOT add bleach or acidic solutions directly to the preparation waste.



Kit Component: Wash Buffer 1 lazard contents Guanidine hydrochloride, 20~50%, CAS-No. 50-01-1 GHS symbol Warning lazard statement(s) Harmful if swallowed. H302 H319 Causes serious eye irritation. ecautionary statem nent(s) Wash ... thoroughly after handling. P264 P280 Wear protective gloves/ protective clothing/ eye protection/ fac protection. IF SWALLOWED: Call a POISON CENTER/ P301 + P312 + P330 doctor if you feel unwell. Rinse mouth

Protocol: Vacuum processing

Please read Important Notes and Additional Materials Required before starting the following steps.

Required hardware

Vacuun manifold for 96-well plate and vaccum source reached to -12 inches \mbox{Hg}

Alternative: If using centrifugation for Elution Step (STEP 8), a centrifuge equiment is required, capable of 5,600~6,000 X g, with a swing-bucket rotor and the adaptor for 96-well plate.

STEP 1. Sample preparation and lysis

- Transfer 200 µl of sample to each well of a Collection Plate (provided, 96-well 2 ml plate; first collection plate).
 If prepared samples are less than 200 µl, adjust sample volume to 200 µl with PBS (not provided).
- Add 400 µl of VNÈ Buffer to each well and mix completely by pipetting.
- · Incubate at room temperature for 10 min.

STEP 2. Adjust binding condition

 \cdot Add 300 μl of AD Buffer (ethanol added) to each well and mix completely by pipetting.

STEP 3. DNA/RNA Binding

- Fix a clean Collection Plate (provided, second collection plate) on the rack of vacuum manifold and cover the manifold lid.
 Place a Filter Plate (provided, 96-Well nucleic acid binding plate) on top of the Collection Plate (second collection plate).
 Transfer the sample mixture to the Filter Plate and discard the
- Collection Plate (first collection plate). Apply vacuum at -12 inches Hg until the wells have emptied.
- Discard the flow-through and return the Filter Plate and the
- Collection Plate to the manifold.

collection rate to the manifold.

STEP 4. Wash the Filter Plate with Wash Buffer 1 \cdot Add 500 μl of Wash Buffer 1 (ethanol added) to each well of the

- Filter Plate. • Apply vacuum at -12 inches Hg until the wells have emptied. • Discard the flow-through and return the Filter Plate and the
- Collection Plate to the manifold.

STEP 5. Wash the Filter Plate with Wash Buffer 2

- \cdot Add 500 μl of Wash Buffer 2 (ethanol added) to each well of the Filter Plate.
- Apply vacuum at -12 inches Hg until the wells have emptied.
 Discard the flow-through and return the Filter Plate and the Collection Plate to the manifold.

STEP 6. Wash the Filter Plate again with Wash Buffer 2

- \cdot Add 500 μl of Wash Buffer 2 (ethanol added) to each well of the Filter Plate.
- Apply vacuum at -12 inches Hg for 10 min.
- Discard the flow-through and return the Collection Plate to the manifold.

STEP 7. Dry the membranes of Filter Plate

- Gently tap the tips of the Filter Plate on a clean paper towel to remove residual liquid.
- Return the Filter Plate to the Collection Plate fixed in the manifold.
- · Apply vacuum for an addition 10 min.
- Discard the flow-through and the Collection Plate (second plate).

STEP 8. DNA/ RNA Elution

- Place the Elution Plate (provided, 96-Well PCR plate) on top of a clean Collection Plate (provided, third collection plate) and fix plates on the rack of manifold. Cover the manifold lid and place the Filter Plate on the Elution Plate. (top: Filter Plate, middle: Elution Plate, bottom: Collection Plate)
- Add 50~75 µl of RNase-free Water to the membrane center of the Filter Plate. Stand for 3 min.
- -- Important Step! For effective elution, make sure that RNase-free water is dispensed on the membrane center and is absorbed completely.
- Important : Do not elute the DNA/RNA using RNase-free water less than suggested volume (<50 µl). It will lower the DNA/RNA yield.
- Close the manifold valve. Turn on the vacuum source to build up a vacuum to -12 inches Hg.
- · Open the manifold valve to apply vacuum to elute DNA/RNA.
- · Seal the Adhesive Film and store the DNA/RNA at -70°C.

Protocol: Centrifuge processing

Please read Important Notes and Additional Materials Required before starting the following steps.

Required hardware

Centrifuge equipment capable of 5,600~6,000 X g with a swingbucket rotor and the adaptor for 96-well plate

STEP 1. Sample preparation and lysis

- Transfer 200 µl of sample to each well of a Collection Plate (provided, 96-well 2 ml plate; first collection plate).
 If prepared samples are less than 200 µl, adjust sample volume to 200 µl with PBS (not provided).
- Add 400 µl of VNE Buffer to each well and mix completely by pipetting.
- · Incubate at room temperature for 10 min.

STEP 2. Adjust binding condition

 Add 300 µl of AD Buffer (ethanol added) to each well and mix completely by pipetting.

STEP 3. DNA/RNA Binding

- Place a Filter Plate (provided, 96-Well nucleic acid binding plate) on a clean Collection Plate (provided, second collection plate).
 Transfer the sample mixture to each well of the Filter Plate and
- discard the Collection Plate (first collection plate).
- Place the plates in a rotor bucket and centrifuge at 5,600~6,000 x g for 2 min.
- Discard the flow-through and return the Filter Plate to the Collection Plate.

STEP 4. Wash the Filter Plate with Wash Buffer 1

- \cdot Add 500 μl of Wash Buffer 1 (ethanol added) to each well of the Filter Plate.
- \cdot Place the combined plate in a rotor bucket and centrifuge at 5,600~6,000 x g for 2 min.
- Discard the flow-through and return the Filter Plate to the Collection Plate.

STEP 5. Wash the Filter Plate with Wash Buffer 2

- \cdot Add 500 μl of Wash Buffer 2 (ethanol added) to each well of the Filter Plate.
- \cdot Place the combined plate in a rotor bucket and centrifuge at 5,600~6,000 x g for 2 min.
- \cdot Discard the flow-through and return the Filter Plate to the Collection Plate.

STEP 6. Wash the Filter Plate again with Wash Buffer 2

- \cdot Add 500 μl of Wash Buffer 2 (ethanol added) to each well of the Filter Plate.
- \cdot Centrifuge at 5,600~6,000 x g for 15 min.
- Discard the flow-through and the Collection Plate (second plate).

STEP 7. Dry the membranes of Filter Plate

• Place the Filter Plate on top of a clean paper towel (not provided) and stand at room temperature for 10 min.

STEP 8. DNA/ RNA Elution

- Place the Elution Plate (provided, 96-Well PCR plate) on top of a clean Collection Plate (provided, third collection plate) then place the Filter Plate on the Elution plate. (top: Filter Plate, middle: Elution Plate, bottom: Collection Plate)
- Add 50~75 µl of RNase-free Water to the membrane center of the Filter Plate. Stand for 3 min.
- Important Step! For effective elution, make sure that RNase-free water is dispensed on the membrane center and is absorbed completely.
- Important : Do not elute the DNA/RNA using RNase-free water less than suggested volume (<50 µl). It will lower the DNA/RNA yield.
- \cdot Place the combined plates in a rotor bucket and centrifuge at 5,600~6,000 x g for 5 min to elute DNA/RNA.
- · Seal the Adhesive Film and store the DNA/RNA at -70°C.

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