

## FAVORGEN\* FavorPrep<sup>TM</sup> PCR Clean Up Micro Kit

-For purification of PCR products or reaction mixtures (concentration and desalination of reaction mixtures)

Cat. No.: FAPC1010 FAPC1013 FAPC1014

For Research Use Only

#### **Kit Contents:**

Cat. No:	FAPC1010	FAPC1013	FAPC1014	
	(4 preps)	(50 preps)	(100 preps)	
MP Buffer Wash Buffer (concentrate) <sup>a</sup> Elution Buffer MP Columns * Collection Tubes User Manual	1.5 ml × 2	30 ml	60 ml	
	1 ml	12.5 ml	22.5 ml	
	0.5 ml	5 ml	5 ml	
	4 pcs	10 × 5 pcs	10 × 10 pcs	
	4 pcs	50 pcs	100 pcs	
	1	1	1	
Preparation of Wash Buffer by adding ethanol (96~100%)				
Ethanol volume for Wash Buffer <sup>a</sup>	4 ml	50 ml	90 ml	

<sup>\*</sup>Store the MP Columns to 4~8°C upon receipt.

## **Specification:**

Principle: Spin column (silica matrix)

DNA Binding capacity of spin column: 5 µg

Sample size: Up to 100 µl of reaction solution

DNA size: 65 bp~10 kbp

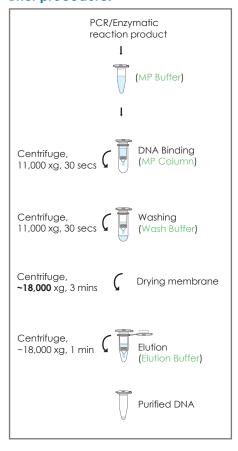
Recovery: 80%~90% for PCR clean up

Operation time: 15 mins
MinimumElution volume: 10 µl

#### **Important Notes:**

- 1. Buffer provided in this kit contain irritants. Wear gloves and lab coat when handling these buffers.
- 2. Add the required volume of ethanol (96~100%) to Wash Buffer before use.
- Centrifugation steps are done by a microcentrifuge capable of the speed at 11,000 ~18,000 xg.

### **Brief procedure:**



#### **Protocol:**

Please Read Important Notes Before Starting Following Steps.

- 1. Transfer 10~100 µl of PCR (or enzymatic product) and 5X volumes of MP Buffer to a 1.5 ml microcentrifuge tube (not provided). Mix by vortexing.
  - -For example, Add 250 µl of MP Buffer to 50 µl of PCR product.
  - -Note: For concentration and purification of DNA from enzymatic reactions, the limits of sample volume and DNA amount are  $100 \, \mu l$  and  $5 \, \mu g$ .
- 2. Place a MP Column to a Collection Tube. Transfer the sample mixture to the MP Column.
- 3. Centrifuge for 1 min then discard the flow-through.
- 4. Add 600 µl of Wash Buffer (ethanol added) to the MP Column. Centrifuge for 1 min then discard the flow-through.
  - -Make sure that ethanol (96~100%) has been added to Wash Buffer at the first open.
- 5. Centrifuge for an additional 3 mins to dry the MP column.
  - -Important step! The residual liquid should be removed completely by this step.
- 6. Place the MP Column into a new 1.5 ml microcentrifuge tube (not provided).
- 7. Add ≥10 µl of Elution Buffer or ddH2O (pH 7.0~8.5) to the membrane center of the MP Column. Stand the MP Column for 2 mins.
  - **-Important step!** For effective elution, make sure that the elution solution is dispensed onto the membrane center and is absorbed completely.
  - -Important: Do not elute the DNA using less than suggested volume (10 µl). It will lower the final yield.
- 8. Centrifuge for 1 min to elute DNA.
  - -The average eluate volume is 9 µl from 10 µl elution buffer volume.

# **Troubleshooting**

Problems	Possible reasons	Solutions	
Low or none recovery of DNA fragment Apply more than 100 µl of PCR product		If PCR product is more than 100 µl, separate it into multiple tubes.	
	Elution of DNA fragment is not efficient	Make sure the pH of Elution Buffer or ddH <sub>2</sub> O is between 7.0~8.5.	
		Make sure that the elution solution has been completely absorbed by the column membrane before centrifugation.	
	The size of DNA fragment is larger than 5 Kb	Preheat the elution solution to 60°C before use.	
Poor perfor- mance in the downstream applications	Salt residue remains in eluted DNA	Wash the column twice with Wash Buffer.	
	Ethanol residue remains in eluted DNA	Do discard the flow-through after washing with Wash Buffer and centrifuge for an additional 3 mins.	