

FavorPrep™ PCR Clean Up Mini Kit

- For purification of PCR products or reaction mixtures
 (concentration and desalination of reaction mixtures)

Cat. No.: FAPC1020
 FAPC1023
 FAPC1025
 FAPC1026

(For Research Use Only)

Kit Contents:

Cat. No:	FAPC1020 (4 preps)	FAPC1023 (50 preps)	FAPC1025 (200 preps)	FAPC1026 (300 preps)
FAPC Buffer	1.5 ml x 2	30 ml	110 ml	180 ml
Wash Buffer (concentrate) [▲]	1 ml	12.5 ml	45 ml	50 ml x 2
Elution Buffer	0.5 ml	5 ml	20 ml	20 ml
FAPC Column	4 pcs	50 pcs	200 pcs	300 pcs
Collection Tube	4 pcs	50 pcs	200 pcs	300 pcs
Elution Tube	4 pcs	50 pcs	200 pcs	300 pcs
User Manual	1	1	1	1
Preparation of Wash Buffer by adding ethanol (96 ~ 100%)				
Ethanol volume for Wash Buffer [▲]	4 ml	50 ml	180 ml	200 ml

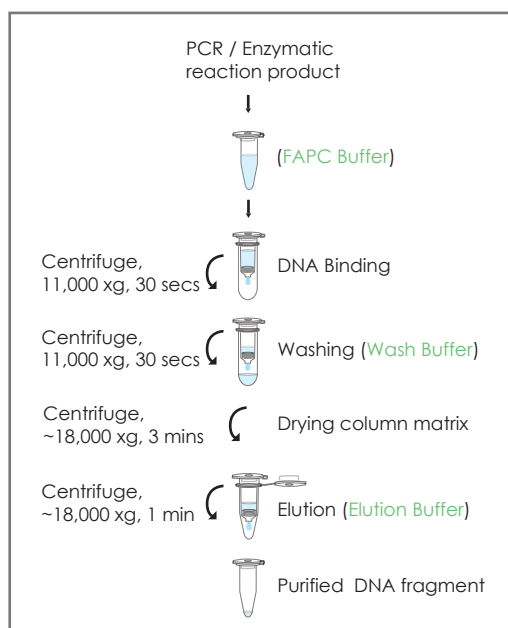
Specification:

Principle: spin column (silica matrix)
 DNA Binding capacity of spin column: 20 µg
 Sample size: up to 100 µl of reaction solution
 DNA size: 65 bp~10 kbp
 Recovery: 85%~95% for PCR clean up
 Operation time: ≤15 mins
 Elution volume: ≥20 µl

Important Notes:

1. Buffer provided in this kit contain irritants. Wear gloves and lab coat when handling these buffer.
2. Add the required volume of ethanol (96~100%) to Wash Buffer before use.
3. For concentration or purification of DNA fragments from enzymatic reactions, the maximum sample volume is 100 µl and the maximum amount of DNA fragments is 5 µg.
4. Centrifugation steps are done by a microcentrifuge capable of the speed at 11,000 ~18,000 xg.

Brief procedure:



General Protocol:

Please Read Important Notes Before Starting Following Steps.

1. Transfer 10~100 μ l of PCR product (excluding oil) to a microcentrifuge tube (not provided) and add 5x volumes of FAPC Buffer, mix well by vortexing.
 - For example, Add 250 μ l of FAPC Buffer to 50 μ l of PCR product.
 - The maximum volume of PCR product is 100 μ l (excluding oil). Do not exceed this limit. If PCR product is more than 100 μ l, separate it into multiple tubes.
2. Place a FAPC column into a Collection Tube.
3. Transfer the sample mixture to the FAPC Column. Centrifuge at 11,000 xg for 30 secs, then discard the flow-through.
4. Add 600 μ l of Wash Buffer (ethanol added) to the FAPC Column. Centrifuge at 11,000 xg for 30 secs, then discard the flow-through.
 - Make sure that ethanol (96-100%) has been added into Wash Buffer when first open.
5. Centrifuge again at full speed (~18,000 xg) for an additional 3 mins to dry the column matrix.
 - **Important step !** The residual liquid should be removed thoroughly on this step.
6. Place the FAPC Column to a Elution Tube. (provided)
7. Add ≥ 20 μ l of Elution Buffer or ddH₂O to the membrane center of the FAPC Column. Stand the FAPC Column for 1 min.
 - **Important step !** For effective elution, make sure that the elution solution is dispensed onto the membrane center and is absorbed completely.
 - **Important :** Do not elute the DNA using less than suggested volume (20 μ l). It will lower the final yield.
8. Centrifuge at full speed (~18,000 xg) for 1 min to elute DNA.

Troubleshooting

Problems	Possible reasons	Solutions
Low or none recovery of DNA fragment	Apply more than 100 μ l of PCR product	If PCR product is more than 100 μ l, separate it into multiple tubes.
	Elution of DNA fragment is not efficient	Make sure the pH of Elution Buffer or ddH ₂ O is between 7.0~8.5.
		Make sure that the elution solution has been completely absorbed by the column membrane before centrifugation.
	The size of DNA fragment is larger than 5 Kb	Preheat the elution solution to 60°C before use.
Poor performance in the downstream applications	Salt residue remains in eluted DNA	Wash the column twice with Wash Buffer.
	Ethanol residue remains in eluted DNA	Do discard the flow-through after washing with Wash Buffer and centrifuge for an additional 3 mins.